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Review Series

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Membranous nephropathy: from models to man

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As recently as 2002, most cases of primary membranous nephropathy (MN), a relatively common cause of nephrotic syndrome in adults, were considered idiopathic. We now recognize that MN is an organ-specific autoimmune disease in which circulating autoantibodies bind to an intrinsic antigen on glomerular podocytes and form deposits of immune complexes in situ in the glomerular capillary walls. Here we define the clinical and pathological features of MN and describe the experimental models that enabled the discovery of the major target antigen, the M-type phospholipase A₂ receptor 1 (PLA₂R). We review the pathophysiology of experimental MN and compare and contrast it with the human disease. We discuss the diagnostic value of serological testing for anti-PLA₂R and tissue staining for the redistributed antigen, and their utility for differentiating between primary and secondary MN, and between recurrent MN after kidney transplant and de novo MN. We end with consideration of how knowledge of the antigen might direct future therapeutic strategies.

Introduction

If one were to search for a condition in which fundamental studies of pathogenesis in animal models have informed major discoveries in the human disease, it would be hard to best the example of membranous nephropathy (MN). Whereas most cases of MN were considered idiopathic as recently as ten years ago, lessons learned from animal models have enabled the discovery of the major target antigen in most adults with MN and defined the causes of less common childhood and rare antenatal cases.

MN, a common cause of the nephrotic syndrome in adults, is an antibody-mediated glomerular disease characterized by the subepithelial formation of immune deposits containing antigen, IgG, and complement components. Sublethal injury to the overlying podocyte leads to cellular simplification and breakdown of the glomerular filtration barrier, causing proteinuria and other manifestations of the nephrotic syndrome. In developed countries, approximately 75% of all MN is primary (or idiopathic) in nature and is considered an organ-specific autoimmune disease, occurring in the absence of any identifying cause or initiating event. The remainder is secondary to conditions such as infection (hepatitis B), systemic autoimmune disease (lupus), medications or exposures (NSAIDs, mercury), and certain malignancies.

Primary MN has a 2:1 male-to-female predominance and a median age of onset in the early 50s, although it may develop anywhere from childhood to advanced ages. Because of its unpredictable natural history, treatment decisions can be challenging. One-third of cases, even those who present with substantial proteinuria, may undergo a spontaneous remission of disease over the course of several years (1). Others may be left with persistent proteinuria but preserved renal function. The most concerning cases involve those in whom high-level proteinuria persists and renal function worsens, often progressing to end-stage renal disease (ESRD), or those that develop complications of the nephrotic syndrome, such as venous thromboembolism. Decisions about when to intervene with potent immunosuppressive therapy are not always straightforward, although clinical guidelines exist (2). In those patients with MN who undergo transplant due to ESRD from MN, the disease may recur in the renal allograft and lead to graft failure.

Pathology, pathophysiology, and clinical correlations

MN was initially named for the thickened (membranous) appearance of the glomerular capillary wall by light microscopy and staged according to the growth of the immune deposits and their incorporation into the expanded glomerular basement membrane (GBM) as seen on EM. We now recognize that the most clinically and immunologically active cases are often those with small subepithelial deposits and no GBM thickening, whereas those with the most advanced stages of GBM expansion may be indolent. Thus, MN is now more typically diagnosed by features on immunofluorescence (IF) and EM. These reveal finely granular immune deposits of IgG (mainly IgG4 in primary MN) in a peripheral capillary loop pattern and electron-dense deposits predominantly or exclusively in a subepithelial location, with effacement of the overlying podocyte foot processes (Figure 1). GBM expansion between and around deposits may or may not be present.

Animal studies have revealed that the subepithelial immune deposits in MN form in situ (Figure 2). Binding of circulating antibodies specific to an intrinsic antigen present on the basal surface of the podocyte is the mechanism at play in most forms of adult MN (see below). Cationic antigens can easily traverse the GBM, become planted in a subepithelial position, and subsequently be targeted by circulating antibodies. This is best exemplified by animal models immunized with cationized BSA (cBSA), in which cBSA binds the negatively charged residues in the GBM and is targeted by circulating anti-BSA antibodies (3). Planted antigens may also be responsible for immune deposits in class V (membranous) lupus nephritis or hepatitis B-associated MN (4, 5). Circulating immune complexes do not generally produce subepithelial deposits and cause MN, but certain physicochemical properties of the complex may enable subendothelial deposits to dissociate and reform under the podocytes (6).

Much of what we know about the pathogenesis of MN derives from observations in the experimental rat model of Heymann nephritis. Studies in the late 1970s established that the subepithelial deposits form in situ when circulating antibodies (resulting from either active or passive immunization of the animal) bind to an intrinsic antigen in the glomerular capillary wall (7, 8). This antigen was subsequently identified as megalin, a member of the LDL receptor family present on the basal surface of rat podocytes (9–11). The binding of circulating anti-megalín antibodies to surface megalín induces capping and shedding of antigen-antibody complexes

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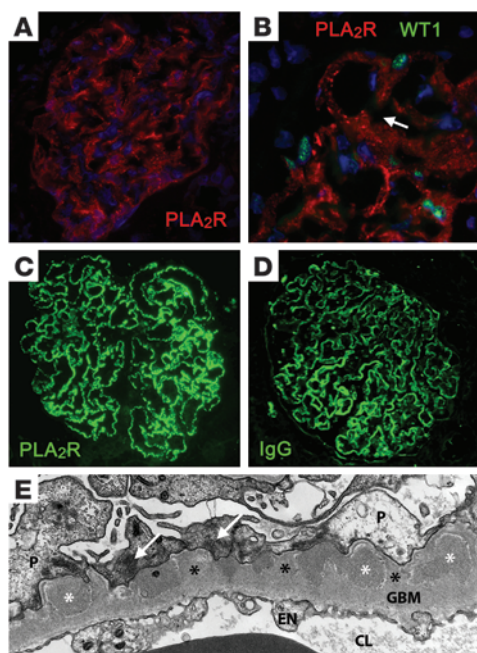


Figure 1

PLA₂R staining in normal and MN glomeruli, and EM of typical subepithelial deposits in MN. (A) IF staining of a normal glomerulus demonstrating PLA₂R expression throughout the podocyte (red). Cell nuclei were counterstained with Hoechst dye (blue). (B) A higher-magnification view of a normal glomerulus shows podocytes, labeled with nuclear WT1 (green), that exhibit PLA₂R (red) staining diffusely throughout the cell body and processes. The portion of the capillary loop covered by mesangium (arrow) did not stain for PLA₂R. In A and B, PLA₂R was stained with a polyclonal anti-PLA₂R antiserum generated in guinea pig, courtesy of G. Lambeau (Institut de Pharmacologie Moléculaire et Cellulaire, CNRS, and Université de Nice-Sophia Antipolis, Valbonne, France). (C and D) PLA₂R staining (green) of a MN kidney biopsy revealed a fine granular capillary loop pattern (C) nearly identical to that of IgG (D). PLA₂R was stained with a commercial anti-PLA₂R antibody generated in rabbit. (E) EM from a patient with primary MN showed electron-dense deposits (white asterisks) in a subepithelial position beneath the podocyte (P) and overlying the GBM. The podocyte exhibited condensation of the actin cytoskeleton and foot process effacement (arrows) and had laid down new ECM material (black asterisks) between the immune deposits. CL, capillary lumen. Original magnification, $\times 400$ (A, C, and D), $\times 630$ (B).

into the underlying GBM. Proteinuria in this model is the result of complement activation by the aggregated complement-fixing antibodies, which are able to overcome local complement inhibitory factors (12). Local generation of the membrane attack complex (MAC; C5b-9) leads to sublethal podocyte injury, resulting in a cascade of structural and functional changes including calcium influx, oxidative injury, production of arachidonic acid metabolites, ER stress, cell cycle dysregulation (13), disruption of the actin cytoskeleton (14), and alterations in the ubiquitin-proteasome system (15, 16). Notably, ubiquitin accumulation and upregulation of autophagy in podocytes has also been described in human MN (17, 18). The loss of the intricate, differentiated cytoskeleton that results from these cellular events induces a simplified cell phenotype with loss and displacement of slit diaphragm structures, leading to a profound non-selective proteinuria (Figure 3). The injured podocyte also produces new ECM that is assembled between and around the immune deposits, giving rise to the characteristic “spikes” and GBM thickening that are particularly well visualized by silver stain.

Several other relevant features of human disease have been identified in the Heymann nephritis model. There is clear evidence that epitope spreading occurs in regions of the megalin protein disparate from the primary epitope (19). The gradual nature of disease resolution is also apparent when a diseased kidney, complete with immune deposits and podocyte foot process effacement, is transplanted into a naive host (20). Proteinuria and podocyte morphology improve over time, but proteinuria can persist indefinitely. Likewise, in human MN complete remission of proteinuria can occur with short duration of immune complex formation, whereas long-standing disease may cause extensive GBM and podocyte remodeling, resulting in incomplete restoration of the filtration barrier and persistent proteinuria despite the absence of ongoing immune injury.

Antigen identification in human disease

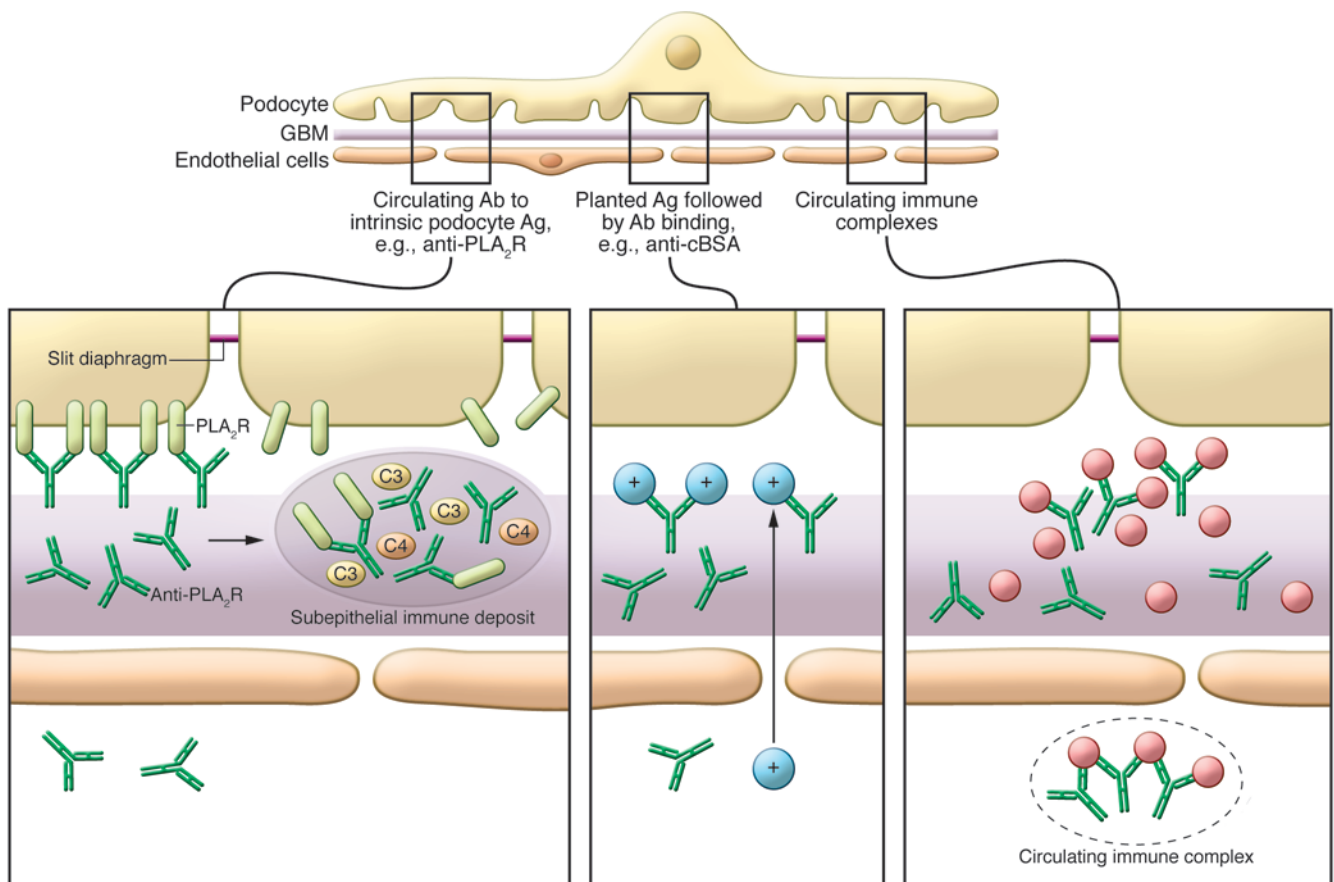
In the past decade, specific proteins have been identified as target antigens in human MN and represent both intrinsic and planted antigens. The first demonstration that circulating antibodies could

target an intrinsic podocyte antigen was identified in a rare case of antenatal MN caused by fetomaternal alloimmunization to neutral endopeptidase (NEP) (21). The mother of the affected child was genetically deficient in NEP and had been alloimmunized during a previous miscarried pregnancy. In the subsequent pregnancy, transplacental passage of anti-NEP antibodies led to in situ antigen-antibody complex formation and complement-mediated podocyte injury in the fetal kidney as a result of alloantibody binding to NEP expressed on the podocytes. Several more cases were subsequently identified, all due to truncating mutations in the maternal gene for NEP (22). Additionally a modified, cationic form of BSA, most likely derived from dietary sources and absorbed intact by the immature intestinal tract of infants, was found to serve as a planted antigen in rare cases of early childhood MN (23).

Both NEP and megalin were ruled out as potential antigens in adult idiopathic MN, but recent evidence indicates that the majority of patients with primary MN have circulating autoantibodies to the M-type phospholipase A₂ receptor 1 (PLA₂R) (24). PLA₂R was identified as the target antigen based on the presence of a high-molecular-weight band in protein extracts from normal human glomeruli that was contingent on intact disulfide bonds. Partial purification of this 180-kDa glycoprotein using lectin chromatography ultimately resulted in the mass spectrometric identification of this band as PLA₂R. Circulating anti-PLA₂R antibodies are detectable in 70%–80% of patients with primary MN and are predominantly but not exclusively IgG4, and all seem to only recognize the protein in the non-reduced state. Consistent with a direct role in pathogenesis, the presence of circulating antibodies to PLA₂R is closely associated with clinical disease activity, the PLA₂R antigen co-localizes with IgG within immune deposits, and IgG reactive with PLA₂R can be specifically eluted from kidney biopsies from patients with MN.

The M-type PLA₂R

PLA₂R is a member of the mannose receptor (MR) family, which also includes the MR, Endo180, and DEC205. It was initially cloned as a receptor for secreted PLA₂ (sPLA₂) and found to have

**Figure 2**

Mechanisms of subepithelial immune deposit formation. Left: Circulating antibodies can target surface-exposed intrinsic podocyte proteins to form in situ immune deposits. In PLA_2R -associated MN, anti- PLA_2R autoantibodies likely bind PLA_2R at the podocyte surface to cause capping and shedding of the antigen-antibody complex into the underlying GBM. Middle: Cationized circulating proteins, such as cBSA, may traverse the GBM and bind beneath the podocyte as planted antigens by virtue of their charge, and also serve as the target for circulating antibodies. Right: There is experimental evidence that circulating immune complexes may initially deposit on the luminal side of the GBM, dissociate, and reform in a subepithelial position. Ag, antigen.

significant expression in human kidney, lung, and placenta (25). However, its exact role in human physiology and in the podocyte has not been fully elucidated. All MR family members have a conserved extracellular structure, with an N-terminal cysteine-rich domain, a fibronectin II-like domain, and eight to ten C-type lectin-like domains. The cytoplasmic domain is short and contains motifs important for the constitutive recycling of these receptors (26). MR family members undergo conformational shifts between a more extended conformation and a compact, folded configuration that may be regulated by pH, oligomerization, and/or ligand binding (27).

Initial cloning showed the highest level of $PLA2R1$ mRNA expression in human kidney (25), although its localization to the human podocyte was not demonstrated until its identification as a target antigen in MN (24). It is expressed throughout the cytoplasm and plasma membrane of the podocyte, but is not expressed by other human glomerular cell types (Figure 1). Gene expression analysis in human kidney supports an expression pattern limited to glomeruli, and in particular, podocytes (28–30). In contrast, this podocyte-restricted pattern in humans is not found in mice (31) or rats (32), where PLA_2R is expressed by glomerular cells other than podocytes.

PLA_2R has been shown to promote replicative senescence in human dermal fibroblasts, as knockdown of human $PLA2R1$ allowed cells to bypass the senescence point and continue proliferating (33). More recent data suggest that PLA_2R may also play a role as a tumor suppressor in mammary epithelium, leading to oncogene-mediated apoptosis via the Jak2 pathway (34). Notably, these effects are independent of s PLA_2 , which further suggests that PLA_2R serves functions other than as a receptor for s PLA_2 . Intriguingly, a PLA_2R molecule devoid of its cytoplasmic domain was equally able to cause cell death, suggesting the presence of an associated molecule for signaling. This is plausible, since another MR family member, Endo180, complexes with the glycoposphatidylinositol-linked urokinase/plasminogen activator receptor and urokinase (35) and appears to play a role in matrix degradation. Several possible functions of PLA_2R in the human podocyte might include a role as a detoxification mechanism for the small (16–20 kDa) s PLA_2 s that are likely filtered through the GBM, a role in maintaining the post-mitotic state of the podocytes, or a role in transcytosis of large-molecular-weight filtered proteins that otherwise might be trapped, analogous to the avian ortholog of PLA_2R , the IgY receptor (or FcRY), that transports IgY from the blood to the yolk sac by transcytosis (36).

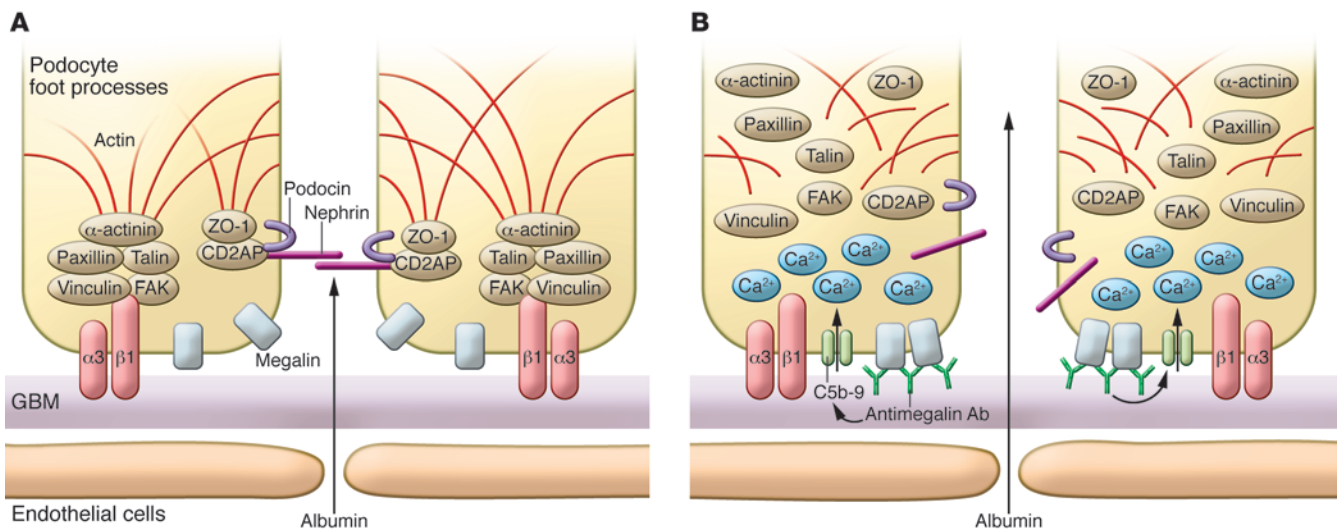


Figure 3
 Mechanisms of podocyte injury and proteinuria in the passive Heymann nephritis (PHN) model of MN. **(A)** The normal podocyte foot process structure is maintained by a well-developed actin cytoskeleton that also serves to anchor the foot processes to the GBM via focal adhesion complexes and cell-matrix adhesion molecules including integrins ($\alpha 3 \beta 1$). The normal filtration slit diaphragm (labeled nephritin) forms a final barrier to albumin permeation and is also linked to the actin cytoskeleton via a complex of proteins. **(B)** In PHN, antibody binding to megalin activates complement leading to assembly and insertion of the MAC (C5b-9). This triggers a cascade of intracellular events (see ref. 90 and the text for details) that contribute to the dissolution of the actin cytoskeleton, which disrupts and displaces the filtration slit diaphragms allowing free passage of albumin into the urine. Collapse of the actin cytoskeleton also affects cell-matrix adhesion and may be the cause of flattening and spreading (effacement) of the podocyte foot processes. Adapted from the *American Journal of Physiology – Renal Physiology* (90).

Anti-PLA₂R autoantibodies

Autoantibodies to PLA₂R (anti-PLA₂R) have emerged as a promising biomarker for the diagnosis and monitoring of immunologic disease activity in primary MN (37, 38). The detection of circulating anti-PLA₂R in a proteinuric subject is highly specific for the presence of primary MN on biopsy (24, 39, 40), and approximately 80% of patients with primary MN will be seropositive for anti-PLA₂R when the disease first becomes clinically apparent (41–45). Small observational and retrospective studies have suggested a temporal relationship between anti-PLA₂R and clinical disease activity. Following the decline and disappearance of anti-PLA₂R (which we term “immunologic remission”), there is a lag over several months before a corresponding clinical remission is seen (46). In this manner, monitoring of anti-PLA₂R may be a quicker and more accurate way of assessing spontaneous remission or efficacy of immunosuppression. Anti-PLA₂R can reappear with disease relapse in the native kidney (46, 47). Similarly, the recurrence of PLA₂R-associated MN in the kidney allograft is also associated with the presence of anti-PLA₂R antibodies (48–51). Early studies that addressed clinical outcome based on anti-PLA₂R titers measured by ELISA suggested that those with the highest levels are less likely to achieve a spontaneous remission (42) and more likely to have worsened kidney function (44), although the numbers on which these conclusions are based are quite small. Future studies will define the optimal use of anti-PLA₂R antibodies.

Several groups are making progress defining the epitope or epitopes within the large PLA₂R molecule that are specifically targeted in disease. Although the major epitope appears conformational in nature and sensitive to reduction of the protein to its primary amino acid sequence (24), reactivity to several discrete linear epitopes has also been described (52).

Tissue staining for PLA₂R in MN

Complementary to the role of circulating anti-PLA₂R as a biomarker, tissue staining for the PLA₂R antigen within immune deposits of MN biopsy specimens has emerged as another method by which to define PLA₂R-associated disease (39, 45, 53, 54). Whereas PLA₂R localizes to the cell body and processes of the normal podocyte, the antigen is preferentially detected in the subepithelial deposits in primary MN, where it co-localizes with IgG (Figure 1). The specificity of this IgG for PLA₂R is suggested by its acid elution from biopsy sections of primary MN patients but not from those of secondary MN or other glomerular disease (24). Similarly, PLA₂R is typically not found in immune deposits in biopsy specimens of patients with secondary forms of MN (39), enabling the use of PLA₂R tissue staining to differentiate primary from secondary MN.

The reason for this apparent change in localization from the cell body to the immune deposits is not entirely clear. One possibility for this enhanced staining in deposits could be that commercial antibody used in the majority of studies detects the aggregated antigen in deposits with greater avidity than it does the molecule in the cell membrane. Dedifferentiation of the podocyte induced by immune injury might result in downregulation of PLA₂R expression. Alternatively, the rate of PLA₂R synthesis may not be sufficient to keep up with the presumed capping and shedding of the molecule into the immune deposits. There was no detectable difference in *PLA2R1* gene expression between MN biopsy samples with or without PLA₂R staining in the deposits (39).

Specificity of anti-PLA₂R for primary versus secondary MN

The recent emergence of PLA₂R-associated disease as an entity has introduced questions about the specificity of these biomarkers for



primary disease. There are rare occurrences of either anti-PLA₂R or tissue PLA₂R in cases that have otherwise been considered secondary. In a cohort of Chinese MN patients, Qin and colleagues describe one case each of HBV- and lupus-associated MN, and three cases of malignancy-associated MN, who were seropositive for anti-PLA₂R (41). Of note, these cases had no features of secondary disease on biopsy and the predominant IgG subclass was IgG4. One interpretation is that, despite the presence of anti-nuclear antibodies, hepatitis B, or malignancy, these patients had primary, PLA₂R-associated disease and another coincidental but causally unrelated disease process. Tissue staining for PLA₂R has not been associated with lupus-associated MN, but has been found in the majority of MN cases associated with sarcoidosis and hepatitis C virus (54, 55), as well some cases felt to be secondary to NSAID use (56). The authors state that there is either a direct association (e.g., these systemic disease processes may activate the immune system to stimulate autoantibody production against PLA₂R) or they may instead be causally unrelated to the primary, PLA₂R-associated disease. We tend to favor the latter explanation, at least in the cases of PLA₂R-associated MN that have positive hepatitis serology or cancer. Interestingly, although MN has been described in several cases of IgG4-related disease (57), this was not associated with PLA₂R deposits, and anti-PLA₂R was not detected in such cases of MN (58–60) or in cases of IgG4-related disease without MN (61).

Genetic associations

Soon after the identification of PLA₂R as the target antigen in primary MN, studies from Korea and Taiwan documented a significant association of MN with SNPs within the *PLA2R1* gene (62, 63). A subsequent unbiased genome-wide association study conducted by a European consortium also revealed a strong genetic association with *PLA2R1*, as well as with *HLA-DQA1* (64). The two most significant SNPs were intronic, but the *PLA2R1* SNP was in strong linkage disequilibrium with the non-synonymous coding SNPs found in the earlier articles. Although the *PLA2R1* and *HLA-DQA1* SNPs were both significantly and independently associated with MN, the odds ratio of MN was almost 80 in individuals who were homozygous for both *HLA-DQA1* and *PLA2R1* variants.

Given the likelihood that PLA₂R, like its MR family relatives, undergoes conformational changes at the cell surface, and due to the fact that the epitope recognized by human autoantibodies is reduction sensitive and thus dependent on secondary or tertiary structure, it has been tempting to speculate that a mutation in *PLA2R1* alters the protein structure, making it a more likely target for autoantibodies. This was directly addressed by exomic sequencing of *PLA2R1* in cases of primary MN, the majority of whom were known to have PLA₂R-associated disease by virtue of circulating antibodies (65). No common coding mutations were found in MN patients with anti-PLA₂R antibodies. It is also revealing that the risk alleles in *PLA2R1* are in fact the major, or more common, alleles in the human population. The strong genetic interaction between the *PLA2R1* and *HLA-DQA1* loci may indicate that a specific HLA molecule is required to present PLA₂R aberrantly or exuberantly to the immune system. A third factor, such as a microbial infection leading to molecular mimicry, may additionally be required for development of disease (66). Although it is unknown whether the primary target engaged by the specific HLA-D receptor is an epitope of PLA₂R or a molecular mimic, the help for B cells to produce IgG4 in MN appears to be driven by Th2 helper cells and their respective cytokines (67).

Genotype-phenotype association has been documented between genetic risk and the presence of circulating anti-PLA₂R. Using a haplotype comprised of two risk alleles in *PLA2R1* and one in *HLA-DQA1*, it was demonstrated that 73% of those homozygous for the high-risk haplotype had circulating anti-PLA₂R, whereas none of those homozygous for the protective haplotype were seropositive for anti-PLA₂R (68). Others have also shown an association between HLA genotype and titers of anti-PLA₂R (44).

Additional antigens

Using a proteomic approach to identify antigens from cultured human podocytes recognized by human MN sera, Ghigerri and colleagues have identified several intracellular enzymes (superoxide dismutase 2; aldose reductase; and α -enolase) also targeted by circulating antibodies (69, 70). These enzymes are not abundantly expressed in the normal glomerulus, but are induced with disease and are thus neoantigens. The prevalence of these autoantibodies is not as high as that for anti-PLA₂R (69), and the temporal relationship of their development or disappearance has not been defined. It is possible that they arise secondarily after an initial insult to the podocyte (perhaps caused by anti-PLA₂R) through the processes of oxidative stress, neoantigen induction, and intermolecular epitope spreading. Whether or not these additional autoantibodies worsen or prolong existing disease, or whether they might be informative as to immunologic duration of disease in non-PLA₂R-associated MN, remains to be seen.

Differences between PLA₂R-associated MN and the paradigm established by Heymann nephritis

Immunohistology of human MN has consistently shown the presence of complement factors within the immune deposits, and complement-mediated podocyte injury has been a cornerstone of the Heymann nephritis experimental model of MN. In the animal model, use of non-complement fixing anti-megalin antibodies, or depletion of terminal complement components, did not cause podocyte injury or proteinuria, despite the presence of IgG-containing immune deposits (71–73). Immune complexes traditionally activate complement via the classical complement pathway, yet primary MN differs from this paradigm view in that the classical complement pathway marker C1q is typically absent. In contrast, the predominant IgG subclass found both within deposits and as the circulating form of anti-PLA₂R is IgG4, a molecule that is not able to bind and activate C1q.

Two non-mutually exclusive possibilities exist to explain this discrepancy. One is based on an observation that the earliest deposits in primary MN have both IgG1 and C1q (74), as well as the fact that there are usually low but detectable levels of the complement-fixing IgG1 and/or IgG3 subclasses of anti-PLA₂R present in the circulation (24). Thus, complement could be activated and sustained at a low level by the classical pathway, even though IgG4 is predominant and may have other immunomodulatory or pathophysiological functions. The second possibility is that IgG4 anti-PLA₂R is able to activate complement via another pathway, such as the lectin pathway. Mannan-binding lectin (MBL) is the initiator of this pathway through the recognition of carbohydrate moieties such as mannose or N-acetyl-glucosamine (GlcNAc) that are not usually exposed on mammalian carbohydrates. MBL has been shown to activate complement in patients with rheumatoid arthritis by binding to a glycan on the Fc portion of IgG that is deficient in terminal galactose, thus exposing GlcNAc (75). Preliminary studies suggest that similar



mechanisms may be at work in the case of IgG4 anti-PLA₂R (76). In either case, initial or weak complement activation by the classical or lectin pathway would be amplified by the alternative pathway.

We should not exclude the possibility that IgG4 anti-PLA₂R may injure podocytes in ways other than direct complement activation. There may be a direct interaction of IgG4 anti-PLA₂R with the PLA₂R molecule in terms of inhibiting (or stimulating) its function, as has been shown for IgG4 autoantibodies to myosin-specific kinase in a subgroup of patients with myasthenia gravis (77). IgG4 has traditionally been viewed as an anti-inflammatory molecule, due to its peculiar characteristics (78). It can exchange arms with other IgG4 molecules with different antigenic specificity and thus lose its bivalency (79). Through this mechanism, IgG4 is limited in its ability to form immune complexes or lattices and may thus downregulate the immune response to a particular antigen. IgG4 also has atypical rheumatoid factor activity and can bind other IgG nonspecifically via Fc-Fc interactions, which may inhibit other complement-fixing antibodies from binding C1q and initiating the complement cascade. However, it seems that PLA₂R-associated MN can exist in the absence of IgG4 anti-PLA₂R. Approximately 5% of cases with circulating anti-PLA₂R lack the IgG4 subclass (42), and an exceptional case of MN caused by monoclonal IgG3κ anti-PLA₂R has recently been described that phenocopies primary MN with the only exception that C1q is present in the immune deposits (50). Further study into the precise pathogenesis and role of the specific anti-PLA₂R subclasses and complement in disease pathogenesis is necessary.

Recurrent MN

Recurrence of primary MN occurs in approximately 40% of cases after kidney transplantation (80). Some of this is subclinical, and may not have otherwise been detected were it not for protocol biopsies. Histologic evidence of recurrent MN can occur as early as one week after transplantation and has been associated with the presence of circulating anti-PLA₂R at the time of transplantation (48, 49). In fact, PLA₂R has been co-localized with IgG in the immune deposits as early as six days after transplantation (51), suggesting that circulating anti-PLA₂R antibodies target the antigen in the allograft. Tiny immune deposits can be detected by EM and, with progressive disease, the size of these deposits increases as does the amount of proteinuria (81). In this way, recurrent disease represents a useful system in which to follow the early roles of circulating anti-PLA₂R in the formation and growth of immune deposits. However, not all patients with circulating anti-PLA₂R at transplantation develop full-blown clinical disease (48). In such cases, genetic differences in the expression and/or conformation of PLA₂R within the allograft may be unfavorable for in situ immune complex formation or the induction therapy and transplant immunosuppression may be sufficient to achieve immunological remission. In those with persistence or re-development of anti-PLA₂R for longer durations, full-blown recurrent MN can occur, often requiring additional immunosuppressive agents such as rituximab (82, 83).

Treatment of MN

Although primary MN remits spontaneously in approximately one-third of cases, immunosuppression is often necessary in the remaining cases due to prolonged severe proteinuria, worsening renal function, or other complications of the nephrotic syndrome such as pulmonary embolism or renal vein thrombosis (84, 85).

The best evidence-based treatment regimens are broadly immunosuppressive, involving alkylating agents or calcineurin inhibitors combined with corticosteroids (2). Due to significant adverse effects of these two primary therapeutic regimens, other immunosuppressive agents have been investigated, often in small randomized trials with a short duration of follow-up, or in observational studies. The newer agents include the anti-B cell agent rituximab, the anti-metabolite mycophenolate mofetil, and ACTH, an immunomodulatory agent and stimulator of endogenous corticosteroid secretion. A more in-depth discussion of these agents can be found in recent reviews or clinical practice guidelines (2, 86, 87).

Although the ultimate goal of treatment in this organ-specific autoimmune disease is termination of the immune response to PLA₂R or other podocyte antigens, the slow decline in circulating antibody titers after treatment (46) places the podocytes at risk for ongoing injury. Thus it is important to devise strategies to interrupt the effector mechanisms of anti-PLA₂R until immunological remission occurs. This might take the form of complement inhibition with newer generations of complement inhibitors or pharmacological interventions to inhibit or activate cellular pathways affected by antibody- and/or complement-mediated podocyte injury (14). Although an early short-term randomized trial with eculizumab, a humanized anti-C5 monoclonal antibody that inhibits the cleavage of C5, failed to show a significant effect on proteinuria or renal function, the dosing regimen may not have achieved effective complement inhibition and patients who received this agent in a 12-month open-label extension trial had a significant decrease of proteinuria (88). Moreover, more recent successful experience with this agent in other complement-mediated kidney diseases and the development of newer complement inhibitors that can be targeted to the site of complement activity are cause for optimism.

The optimal and least toxic therapy for such a disease as MN would be the targeted deletion of B cells and plasma cells producing pathogenic antibodies. While it may still be quite some time before such targeted therapy is available for clinical use, a mechanism has recently been identified to eliminate antigen-specific B cells using nanoparticles containing the antigen and a carbohydrate ligand for SigLec, which confers an inhibitory signal to the B cell receptor when complexed with antigen (89). Once the primary PLA₂R epitope is defined, other specific interventions, such as restoring tolerance by oral immunization or the development of antibody traps or decoys, may become feasible. The potential for epitope spreading to other parts of the PLA₂R molecule during disease progression, however, may make such strategies more challenging.

Future understanding of disease pathogenesis in PLA₂R-associated MN and identification of other target antigens in idiopathic primary MN will continue to evolve and improve our diagnosis, monitoring, and therapy of this fascinating yet challenging disease.

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1. Polanco N, et al. Spontaneous remission of nephrotic syndrome in idiopathic membranous nephropathy. *J Am Soc Nephrol*. 2010;21(4):697–704.
2. Kidney Disease: Improving Global Outcomes (KDIGO) Glomerulonephritis Work Group. KDIGO Clinical Practice Guideline for Glomerulonephritis. *Kidney Int Suppl*. 2012;2(2):139–274.
3. Border WA, Ward HJ, Kamil ES, Cohen AH. Induction of membranous nephropathy in rabbits by administration of an exogenous cationic antigen. *J Clin Invest*. 1982;69(2):451–461.
4. Schmiedeke TM, Stöckl FW, Weber R, Sugisaki Y, Batsford SR, Vogt A. Histones have high affinity for the glomerular basement membrane. Relevance for immune complex formation in lupus nephritis. *J Exp Med*. 1989;169(6):1879–1894.
5. Johnson RJ, Couser WG. Hepatitis B infection and renal disease: clinical, immunopathogenetic and therapeutic considerations. *Kidney Int*. 1990;37(2):663–676.
6. Fujigaki Y, Nagase M, Honda N. Intraglomerular basement membrane translocation of immune complex (IC) in the development of passive in situ IC nephritis of rats. *Am J Pathol*. 1993;142(3):831–842.
7. Couser WG, Steinmuller DR, Stilmant MM, Salant DJ, Lowenstein LM. Experimental glomerulonephritis in the isolated perfused rat kidney. *J Clin Invest*. 1978;62(6):1275–1287.
8. Van Damme BJ, Fleuren GJ, Bakker WW, Vernier RL, Hoedemaeker PJ. Experimental glomerulonephritis in the rat induced by antibodies directed against tubular antigens. V. Fixed glomerular antigens in the pathogenesis of heterologous immune complex glomerulonephritis. *Lab Invest*. 1978;38(4):502–510.
9. Farquhar MG, Saito A, Kerjaschki D, Orlando RA. The Heymann nephritis antigenic complex: megalin (gp330) and RAP [editorial]. *J Am Soc Nephrol*. 1995;6(1):35–47.
10. Kerjaschki D, Farquhar MG. The pathogenic antigen of Heymann nephritis is a membrane glycoprotein of the renal proximal tubule brush border. *Proc Natl Acad Sci U S A*. 1982;79(18):5557–5561.
11. Kerjaschki D, Farquhar MG. Immunocytochemical localization of the Heymann nephritis antigen (GP330) in glomerular epithelial cells of normal Lewis rats. *J Exp Med*. 1983;157(2):667–686.
12. Schiller B, He C, Salant DJ, Lim A, Alexander JJ, Quigg RJ. Inhibition of complement regulation is key to the pathogenesis of active Heymann nephritis. *J Exp Med*. 1998;188(7):1353–1358.
13. Shankland SJ, et al. Cyclin kinase inhibitors are increased during experimental membranous nephropathy: potential role in limiting glomerular epithelial cell proliferation in vivo. *Kidney Int*. 1997;52(2):404–413.
14. Takano T, Elimam H, Cybulsky AV. Complement-mediated cellular injury. *Semin Nephrol*. 2013;33(6):586–601.
15. Meyer-Schwesinger C, et al. A new role for the neuronal ubiquitin C-terminal hydrolase-L1 (UCH-L1) in podocyte process formation and podocyte injury in human glomerulopathies. *J Pathol*. 2009;217(3):452–464.
16. Kitzler TM, Papillon J, Guillemette J, Wing SS, Cybulsky AV. Complement modulates the function of the ubiquitin-proteasome system and endoplasmic reticulum-associated degradation in glomerular epithelial cells. *Biochim Biophys Acta*. 2012;1823(5):1007–1016.
17. Meyer-Schwesinger C, et al. Ubiquitin C-terminal hydrolase-11 activity induces polyubiquitin accumulation in podocytes and increases proteinuria in rat membranous nephropathy. *Am J Pathol*. 2011;178(5):2044–2057.
18. Hartleben B, et al. Autophagy influences glomerular disease susceptibility and maintains podocyte homeostasis in aging mice. *J Clin Invest*. 2010;120(4):1084–1096.
19. Shah P, Tramontano A, Makker SP. Intramolecular epitope spreading in Heymann nephritis. *J Am Soc Nephrol*. 2007;18(12):3060–3066.
20. Makker SP, Kanalas JJ. Course of transplanted Heymann nephritis kidney in normal host. Implications for mechanism of proteinuria in membranous glomerulonephropathy. *J Immunol*. 1989;142(10):3406–3410.
21. Debiec H, et al. Antenatal membranous glomerulonephritis due to anti-neutral endopeptidase antibodies. *N Engl J Med*. 2002;346(26):2053–2060.
22. Debiec H, et al. Role of truncating mutations in MME gene in fetomaternal alloimmunisation and antenatal glomerulopathies. *Lancet*. 2004;364(9441):1252–1259.
23. Debiec H, et al. Early-childhood membranous nephropathy due to cationic bovine serum albumin. *N Engl J Med*. 2011;364(22):2101–2110.
24. Beck LH Jr, et al. M-type phospholipase A2 receptor as target antigen in idiopathic membranous nephropathy. *N Engl J Med*. 2009;361(1):11–21.
25. Ancian P, Lambeau G, Mattei MG, Lazdunski M. The human 180-kDa receptor for secretory phospholipases A2. Molecular cloning, identification of a secreted soluble form, expression, and chromosomal localization. *J Biol Chem*. 1995;270(15):8963–8970.
26. Zvaritch E, Lambeau G, Lazdunski M. Endocytic properties of the M-type 180-kDa receptor for secretory phospholipases A2. *J Biol Chem*. 1996;271(1):250–257.
27. Llorca O. Extended and bent conformations of the mannose receptor family. *Cell Mol Life Sci*. 2008;65(9):1302–1310.
28. Nyström J, Fierlbeck W, Granqvist A, Kulak SC, Ballermann BJ. A human glomerular SAGE transcriptome database. *BMC Nephrol*. 2009;10:13.
29. Lindemeyer MT, et al. Systematic analysis of a novel human renal glomerulus-enriched gene expression dataset. *PLoS One*. 2010;5(7):e11545.
30. Ju W, et al. Defining cell-type specificity at the transcriptional level in human disease. *Genome Res*. 2013;23(11):1862–1873.
31. Boerries M, et al. Molecular fingerprinting of the podocyte reveals novel gene and protein regulatory networks. *Kidney Int*. 2013;83(6):1052–1064.
32. Beck S, et al. Upregulation of group IB secreted phospholipase A(2) and its M-type receptor in rat ANTI-THY-1 glomerulonephritis. *Kidney Int*. 2006;70(7):1251–1260.
33. Augert A, Payre C, de Launoit Y, Gil J, Lambeau G, Bernard D. The M-type receptor PLA2R regulates senescence through the p53 pathway. *EMBO Rep*. 2009;10(3):271–277.
34. Vindrieux D, et al. PLA2R1 mediates tumor suppression by activating JAK2. *Cancer Res*. 2013;73(20):6334–6345.
35. Behrendt N. The urokinase receptor (uPAR) and the uPAR-associated protein (uPARAP/Endo180): membrane proteins engaged in matrix turnover during tissue remodeling. *Biol Chem*. 2004;285(2):103–136.
36. Tesar DB, Cheung EJ, Bjorkman PJ. The chicken yolk sac IgY receptor, a mammalian mannose receptor family member, transcytoses IgY across polarized epithelial cells. *Mol Biol Cell*. 2008;19(4):1587–1593.
37. Hofstra JM, Wetzels JF. Anti-PLA(2)R antibodies in membranous nephropathy: ready for routine clinical practice? *Neth J Med*. 2012;70(3):109–113.
38. Schlumberger W, et al. Differential diagnosis of membranous nephropathy with autoantibodies to phospholipase A2 receptor 1. *Autoimmun Rev*. 2014;13(2):108–113.
39. Hoxha E, et al. Enhanced expression of the M-type phospholipase A2 receptor in glomeruli correlates with serum receptor antibodies in primary membranous nephropathy. *Kidney Int*. 2012;82(7):797–804.
40. Hoxha E, et al. An immunofluorescence test for phospholipase-A2-receptor antibodies and its clinical usefulness in patients with membranous glomerulonephritis. *Nephrol Dial Transplant*. 2011;26(8):2526–2532.
41. Qin W, et al. Anti-phospholipase A2 receptor antibody in membranous nephropathy. *J Am Soc Nephrol*. 2011;22(6):1137–1143.
42. Hofstra JM, et al. Antiphospholipase A2 receptor antibody titer and subclass in idiopathic membranous nephropathy. *J Am Soc Nephrol*. 2012;23(10):1735–1743.
43. Oh YJ, Yang SH, Kim DK, Kang SW, Kim YS. Autoantibodies against phospholipase A2 receptor in Korean patients with membranous nephropathy. *PLoS One*. 2013;8(4):e62151.
44. Kanigicherla D, et al. Anti-PLA2R antibodies measured by ELISA predict long-term outcome in a prevalent population of patients with idiopathic membranous nephropathy. *Kidney Int*. 2013;83(5):940–948.
45. Svobodova B, Honsova E, Ronco P, Tesar V, Debiec H. Kidney biopsy is a sensitive tool for retrospective diagnosis of PLA2R-related membranous nephropathy. *Nephrol Dial Transplant*. 2013;28(7):1839–1844.
46. Beck LH Jr, et al. Rituximab-induced depletion of anti-PLA2R autoantibodies predicts response in membranous nephropathy. *J Am Soc Nephrol*. 2011;22(8):1543–1550.
47. Hofstra JM, Beck LH Jr, Beck DM, Wetzels JF, Salant DJ. Anti-phospholipase A2 receptor antibodies correlate with clinical status in idiopathic membranous nephropathy. *Clin J Am Soc Nephrol*. 2011;6(6):1286–1291.
48. Debiec H, et al. Autoantibodies specific for the phospholipase A2 receptor in recurrent and De Novo membranous nephropathy. *Am J Transplant*. 2011;11(10):2144–2152.
49. Stahl R, Hoxha E, Fechner K. PLA2R autoantibodies and recurrent membranous nephropathy after transplantation. *N Engl J Med*. 2010;363(5):496–498.
50. Debiec H, et al. Recurrent membranous nephropathy in an allograft caused by IgG3κ targeting the PLA2 receptor. *J Am Soc Nephrol*. 2012;23(12):1949–1954.
51. Blosser CD, Ayalon R, Nair R, Thomas C, Beck LH Jr. Very early recurrence of anti-phospholipase A2 receptor-positive membranous nephropathy after transplantation. *Am J Transplant*. 2012;12(6):1637–1642.
52. Behner A, et al. An anti-phospholipase A2 receptor quantitative immunoassay and epitope analysis in membranous nephropathy reveals different antigenic domains of the receptor. *PLoS One*. 2013;8(4):e61669.
53. Debiec H, Ronco P. PLA2R autoantibodies and PLA2R glomerular deposits in membranous nephropathy. *N Engl J Med*. 2011;364(7):689–690.
54. Larsen CP, Messias NC, Silva FG, Messias E, Walker PD. Determination of primary versus secondary membranous glomerulopathy utilizing phospholipase A2 receptor staining in renal biopsies. *Mod Pathol*. 2013;26(5):709–715.
55. Knehtl M, et al. A case of phospholipase A receptor-positive membranous nephropathy preceding sarcoid-associated granulomatous tubulointerstitial nephritis. *Am J Kidney Dis*. 2011;57(1):140–143.
56. Nawaz FA, Larsen CP, Troxell ML. Membranous nephropathy and nonsteroidal anti-inflammatory agents. *Am J Kidney Dis*. 2013;62(5):1012–1017.
57. Alexander MP, et al. Membranous glomerulonephritis is a manifestation of IgG4-related disease. *Kidney Int*. 2013;83(3):455–462.
58. Cravedi P, et al. Salant DJ, Benigni A, et al. Membranous nephropathy associated with IgG4-related disease. *Am J Kidney Dis*. 2011;58(2):272–275.
59. Fervenza FC, Downer G, Beck LH Jr, Sethi S. IgG4-related tubulointerstitial nephritis with membranous nephropathy. *Am J Kidney Dis*. 2011;58(2):320–324.



60. Li XL, et al. IgG4-related membranous nephropathy with high blood low urine IgG4/IgG ratio: a case report review of the literature. *Clin Rheumatol*. 2014; 33(1):145–148.

61. Khosroshahi A, Ayalon R, Beck LH Jr, Salant DJ, Bloch DB, Stone JH. IgG4-related disease is not associated with antibody to the phospholipase A2 receptor. *Int J Rheumatol*. 2012;2012:139409.

62. Liu YH, et al. Association of phospholipase A2 receptor 1 polymorphisms with idiopathic membranous nephropathy in Chinese patients in Taiwan. *J Biomed Sci*. 2010;17(1):81.

63. Kim S, et al. Single nucleotide polymorphisms in the phospholipase A2 receptor gene are associated with genetic susceptibility to idiopathic membranous nephropathy. *Nephron Clin Pract*. 2010; 117(3):c253–c258.

64. Stanescu HC, et al. Risk HLA-DQA1 and PLA(2)R1 alleles in idiopathic membranous nephropathy. *N Engl J Med*. 2011;364(7):616–626.

65. Coenen MJ, et al. Phospholipase A2 receptor (PLA2R1) sequence variants in idiopathic membranous nephropathy. *J Am Soc Nephrol*. 2013; 24(4):677–683.

66. Salant DJ. Genetic variants in membranous nephropathy: perhaps a perfect storm rather than a straightforward conformeropathy? *J Am Soc Nephrol*. 2013;24(4):525–528.

67. Kuroki A, Iyoda M, Shibata T, Sugisaki T. Th2 cytokines increase and stimulate B cells to produce IgG4 in idiopathic membranous nephropathy. *Kidney Int*. 2005;68(1):302–310.

68. Lv J, et al. Interaction between PLA2R1 and HLA-DQA1 variants associates with anti-PLA2R antibodies and membranous nephropathy. *J Am Soc Nephrol*. 2013;24(8):1323–1329.

69. Murtas C, et al. Coexistence of different circulating anti-podocyte antibodies in membranous nephropathy. *Clin J Am Soc Nephrol*. 2012;7(9):1394–1400.

70. Prunotto M, et al. Autoimmunity in membranous nephropathy targets aldose reductase and SOD2. *J Am Soc Nephrol*. 2010;21(3):507–519.

71. Salant DJ, Belok S, Madaio MP, Couser WG. A new role for complement in experimental membranous nephropathy in rats. *J Clin Invest*. 1980; 66(6):1339–1350.

72. Baker PJ, Ochi RF, Schulze M, Johnson RJ, Campbell C, Couser WG. Depletion of C6 prevents development of proteinuria in experimental membranous nephropathy in rats. *Am J Pathol*. 1989; 135(1):185–194.

73. Cybulsky AV, Rennke HG, Feintzeig ID, Salant DJ. Complement-induced glomerular epithelial cell injury. Role of the membrane attack complex in rat membranous nephropathy. *J Clin Invest*. 1986; 77(4):1096–1107.

74. Huang CC, et al. IgG subclass staining in renal biopsies with membranous glomerulonephritis indicates subclass switch during disease progression. *Mod Pathol*. 2013;26(6):799–805.

75. Malhotra R, Wormald MR, Rudd PM, Fischer PB, Dwek RA, Sim RB. Glycosylation changes of IgG associated with rheumatoid arthritis can activate complement via the mannose-binding protein. *Nat Med*. 1995;1(3):237–243.

76. Ma H, Beck LH, Salant DJ. Membranous nephropathy-associated anti-phospholipase A2 receptor IgG4 autoantibodies activate the lectin complement pathway [abstract]. *J Am Soc Nephrol*. 2011;22:62A.

77. Klooster R, et al. Muscle-specific kinase myasthenia gravis IgG4 autoantibodies cause severe neuromuscular junction dysfunction in mice. *Brain*. 2012; 135(pt 4):1081–1101.

78. Aalberse RC, Stapel SO, Schuurman J, Rispens T. Immunoglobulin G4: an odd antibody. *Clin Exp Allergy*. 2009;39(4):469–477.

79. van der Neut Kolfschoten M, et al. Anti-inflammatory activity of human IgG4 antibodies by dynamic Fab arm exchange. *Science*. 2007;317(5844):1554–1557.

80. Dabade TS, Grande JP, Norby SM, Fervenza FC, Cosio FG. Recurrent idiopathic membranous nephropathy after kidney transplantation: a surveillance biopsy study. *Am J Transplant*. 2008; 8(6):1318–1322.

81. Rodriguez EF, et al. The pathology and clinical features of early recurrent membranous glomerulonephritis. *Am J Transplant*. 2012;12(4):1029–1038.

82. El-Zoghby ZM, Grande JP, Fraile MG, Norby SM, Fervenza FC, Cosio FG. Recurrent idiopathic membranous nephropathy: early diagnosis by protocol biopsies and treatment with anti-CD20 monoclonal antibodies. *Am J Transplant*. 2009; 9(12):2800–2807.

83. Sprangers B, et al. Beneficial effect of rituximab in the treatment of recurrent idiopathic membranous nephropathy after kidney transplantation. *Clin J Am Soc Nephrol*. 2010;5(5):790–797.

84. Barbour SJ, et al. Disease-specific risk of venous thromboembolic events is increased in idiopathic glomerulonephritis. *Kidney Int*. 2012;81(2):190–195.

85. Lionaki S, et al. Venous thromboembolism in patients with membranous nephropathy. *Clin J Am Soc Nephrol*. 2012;7(1):43–51.

86. Waldman M, Austin HA. Treatment of idiopathic membranous nephropathy. *J Am Soc Nephrol*. 2012; 23(10):1617–1630.

87. Hofstra JM, Fervenza FC, Wetzels JF. Treatment of idiopathic membranous nephropathy. *Nat Rev Nephrol*. 2013;9(8):443–458.

88. Cunningham PN, Quigg RJ. Contrasting roles of complement activation and its regulation in membranous nephropathy. *J Am Soc Nephrol*. 2005; 16(5):1214–1222.

89. Nitschke L. Suppressing the antibody response with Siglec ligands. *N Engl J Med*. 2013; 369(14):1373–1374.

90. Cybulsky AV, Quigg RJ, Salant DJ. Experimental membranous nephropathy redux. *Am J Physiol Renal Physiol*. 2005;289(4):F660–F671.